

Review Article

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Challenges and Perspectives on Plasmid Curing, Medicinal and Pharmacological Traits of Plumbago Zeylanica (Chitraka): A Review

Rajashree B. Patwardhan*¹

*¹ Associate Professor, Department of Microbiology, Haribhai V. Desai College of Arts, Science and Commerce Affiliated to Savitribai Phule Pune University, Maharashtra State, Pune 411002, India.

Abstract: From ancient times of vedas, charaksamhita and sushrutsamhita, to modern day developments and research in medicine, the medicinal importance of Chitraka (Plumbago zeylanica) as a wonderful Indian remedy has been upheld through the test of time. Chitraka is used in ayurveda for relief from many ailments, especially digestive disorders, bronchitis, diseases of liver, leucoderma, inflammation, piles, itching, laryngitis, rheumatism, diseases of spleen, many skin disorders etc. The root extracts of P. zeylanica have been incorporated in various Indian indigenous ayurvedic drug formulations. This paper reviews various aspects of Chitraka like different pharmacological activities, medicinal properties including wound healing, antioxidant, antiulcer, anticancer, leishmanicidal, antifertility, antimalarial, antidiabetic, hypolipidemic, trypanocidal, antibacterial, antifungal, antiviral, anti-inflammatory, antimutagenic, anti-allergic, larvicidal, insecticidal and anxiolytic activities. P. zeylanica plant contains naphthoquinones, flavonoids, terpenes, alkaloids, glycosides, steroids, triterpenoids, tannins, phenolic compounds, glucopyranoside, sitosterol saponins, coumarins, carbohydrates, fixed oils, fats and proteins having a wide variety of bioactivities. The important compound responsible for bioactivity is plumbagin which is chemically 5-hydroxy-2-methyl-1,4-naphthoquinone. Studies of P. zeylanica roots resulted in identification of plumbagin and lawsone as an active principle exhibiting the plasmid elimination activity. Due to the toxicity of chemical curing agents like acridine orange or ethidium bromide, there is a constant need of developing novel curing agents which are more effective and at the same time their nontoxic nature. Bacterial strains resistant to multiple antibiotics have emerged to which the invention of new antibiotics has failed to match up. The effects of antibiotic resistance are serious with mortality and morbidity constantly on the rise. Therefore P. zeylanica root extracts containing lawsone and plumbagin would have great potential as drugs of choice in the treatment of antibiotic resistant bacterial strains. The already ineffective antibiotic therapy can be made effective by converting antibiotic resistant bacteria into sensitive ones. The present review for the first time depicts the use of P. zeylanica as antimicrobial and plasmid curing agent in medicinal formulations and it is a novel approach towards the spread of antibiotic resistance especially in the hospital environment.

Keywords: Plumbago zeylanica, Plasmid curing, Bioactive compounds, Plumbagin, Lawsone

*Corresponding Author

Rajashree B. Patwardhan , Associate Professor, Department of Microbiology, Haribhai V. Desai College of Arts, Science and Commerce Affiliated to Savitribai Phule PuneUniversity, Maharashtra State, Pune 411002, India.



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I. INTRODUCTION

There is a continuing search for new antimicrobial compounds from other sources including plants as they are known to produce diverse bioactive substances of chemotherapeutic value¹. The most important of these bioactive compounds are alkaloids, flavonoids, tannins, and phonetic compounds². According to the World Health Organization medicinal plants would be the best source to obtain a variety of drugs³. These are not only used for primary health care not just in rural areas in developing countries, but also in developed countries as well where modern medicines are predominantly used⁴. Such plants should be investigated to understand their properties, safety, and efficiency⁵. Microorganisms have developed resistance to many antibiotics which has created immense clinical problems in treatment of infectious diseases. Clinically most important resistance to antibiotics is the result of plasmid encoded genes⁶. Presence of antibiotic resistance genes on bacterial plasmids and transposons has further helped in transmission and spread of drug resistance among pathogenic bacteria like E. coli, Shigella, Salmonella, Acinetobacter, Staphylococcus, S. pneumoniae and M. tuberculosis ^{7,8,9}. Secondary metabolites produced by plants constitute a source of bioactive substances and now a day's scientific interest has increased due to search for new drugs from plant origin. Hence, more studies pertaining to the use of plants as therapeutic agents should be emphasized, especially those related to control of antibiotic resistant microbes. This has encouraged research into screening of plants for antimicrobial activities. Chitraka i.e. Plumbago is known as "Vanaushadhi plant" (medicinal plant from forest) since ancient times in India and it is interesting to note that its reference is found even in Vedas¹⁰. Even thereafter we find its reference in ancient "Charaksamhita", as a plant used in various medicinal applications to ensure overall health¹¹. It is also referred by "Sushrut"¹². Chitraka literally means "agni" i.e. fire which has capacity to "burn" the disorders¹³. Charaksamhita gives details of various mixtures and medical preparations in which products of "Chitraka" are used to cure various diseases and disorders¹¹. The importance of Chitraka is mentioned in Charaksamhita Adhyaya 15. Charaksamhita explains as to how since ancient times, Chitraka i.e. Plumbago zeylanica has been used for ayurvedic treatment, due to its medicinal properties and effects which were experienced since then. It is used in traditional system of Indian medicine against several ailments including diarrhoea, leprosy, digestive disorders, bronchitis, diseases of liver, leucoderma, inflammation, piles, itching, laryngitis, rheumatism, diseases of spleen and many skin disorders¹⁴. Chitraka stimulates digestive power and helps to accelerate the appetite¹⁵. Biological activities of

crude extracts and active constituents of this plant reported so far include antimicrobial, antimutagenic, antitumor and radio-modifying properties¹⁶. The pulped roots or aerial parts are abortifacient, while powdered bark, root or leaves are used to treat gonorrhea, syphilis, tuberculosis, rheumatic pain, swellings, wound healing¹⁷ dyspepsia, piles, diarrhoea, skin diseases, leprosy and also reported to possess antibacterial, antifungal properties¹⁸. Even after the advent of modern branches of science like botany, ethnobotany, microbiology, pharmacognosy, various properties and effects of Chitraka have been studied extensively and it is found to be more and more useful even in modern day medicine. Plumbago roots contain naphthoquinones, the chemical compounds having a wide variety of bioactivities^{19,20}. With the revitalisation of herbal plants across the world, P. zeylanica is widely used for commercial preparation of drugs due to its biological activities²¹. This review is an effort to bring together all the properties and effects of *P. zeylanica* root extracts including plasmid curing and research thereon and expanding horizons thereof in treating patients against various health problems arising out of resistance of bacteria.

2. CHEMICAL CONSTITUENTS OF PLUMBAGO ROOTS

Due to the remarkable traditional medicinal properties P. zeylanica roots have been extensively screened for their chemical constituents. Major compounds isolated from these plants are naphthoquinones, flavonoids, terpenes, and sterols^{22,23,24}. The 1,4-naphthoquinones are important metabolites of P. zeylanica. These naphthoquinones are derived mostly by substitution or oligomerisation of monomer, plumbagin. The plant contains a number of naphthoquinone derivatives consisting of monomers, dimers, and trimers²⁰. Terpenes include lupeol, lupeol acetate, friedelinol and lupanone. Sterols are β -sitosterol, sitosterone, stigmasterol and stigmasterol acetate. Other chemical constituents include vanillic acid, plumbagic acid, glucose, unidentified tannin, and unidentified glycoside. Free amino acids of P. zeylanica include aspartic acid, tryptophane, tyrosine, threonine, histidine, glycine, hydroxyproline, alanine and methionine²⁵. Nine compounds were isolated from aerial parts of P. zeylanica which includes plumbagin (I), isoshinanolone (II), plumbagic acid (III), betasitosterol (IV), 4-hydroxybenzaldehyde (V), trans-cinnamic (VI), vanillic acid (VII), 2, 5-dimethyl-7acid hydroxychromone (VIII), indole-3-carboxaldehyde (IX)^{26,27}. In the research by Patwardhan et al Lawsone was isolated for the first time from the roots of P. zeylanica 28 .

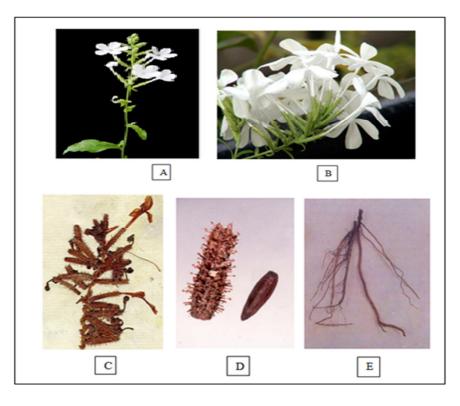


Figure. I Plumbago zeylanica plant with different parts

Figure Ligand

A.and B: Plumbago zeylanica branch with white flowers:
C: P. zeylanica flowers with glandular and elongated spikes.
D: P. zeylanica seed, length 5m
E: Roots of P. zeylanica

3. BROAD SPECTRUM MEDICINAL PROPERTIES OF PLUMBAGO ZEYLANICA

The plant has been used in Indian medicine since the period of Charaka in treatment against inflammations, pile, krimi (worms) and kushtha (various skin diseases)^{29,30,31}. Ρ zeylanica and other Plumbago species are widely used in several oriental systems of medicine in India, China and Eastern countries like Taiwan, Korea, and Malaysia ^{23,32}. Roots and root bark are bitter, stomachic, carminative, and astringent to bowels^{33,34}, antihelminthic, cure intestinal troubles, dysentery, inflammation, piles, bronchitis, itching, and diseases of liver ¹⁴. Root extract is used as a laxative, expectorant, tonic, good appetizer, useful in laryngitis, rheumatism³⁵, diseases of spleen, ringworm, and scabies ³⁴. Paste of root with milk, vinegar or salt and water, is applied to open abscesses, leprosy, other skin diseases externally³⁶. The renoprotective effect of hydroalcoholic extract P. zeylanica was observed in Swiss albino mice³⁷.Plumbagin from P. zeylanica stimulates the central nervous system in small doses. It has well marked antiseptic properties³⁴. Plumbagin was found to exhibit fairly good results in early leucoderma and baldness³⁸. Plumbagin and its dimmer 3,3'-biplumbagin have been used in treatment of leishmaniasis^{39,40}. The important compound responsible for bioactivity is plumbagin, chemically 5-hydroxy-2-methyl-1,4-naphthoquinone. This was studied for its effect on development of antibiotic resistance using antibiotic sensitive strains of E. coli and S. aureus⁴¹. Crude extract of P. zeylanica containing naphthoquinones was found effective as antimicrobial and plasmid curing agent⁴². P. zeylanica root powder displayed estrogenic properties. Three endopeptidases (cathepsin D, renin and chymotrypsin) were

studied in the uterus of albino rats after administration of *P. zeylanica* root powder. The changes were compared with effects induced by 17- β -estradiol in the same experimental conditions. Physiological activities of *P. zeylanica* root powder and 17-beta-estradiol mediated and modified in presence of ovaries. Presence of one or both ovaries modified the activities of enzymes. The results confirmed the estrogenic properties of *P. zeylanica* root powder ^{43,44}. The details of medicinal properties reported includes

3.1 Wound Healing Activity

Herbal extract of *P. zeylanica* was used in combination with *Rubia cordifolia, Centella asiatica, Terminalia belerica, Withania somnifera* and wound healing activity was evaluated in albino rats. The drug was used in ointment dosage form and then compared with a marketed formulation (Soframycin cream) as reference drug. The herbal drug combination has been observed to promote healing of wounds in animals^{45,46}. Wound healing activity of methanolic extract of *P. zeylanica* roots have been reported in wistar albino rats. This study explored the wound healing action of ethanolic root extract of *P. zeylanica* in wistar rats and discovered that the activity is due to to the presence of phytochemicals such as terpenoids, alkaloids, flavonoids, saponins etc. and these compounds are responsible for the wound healing activity of the *P. zeylanica* plant.

3.2 Antioxidant Activity

Extracts of *P. zeylanica* and its active ingredient plumbagin have substantial antioxidant capabilities⁴⁷. CapsHT2, a

polyherbal preparation which consist of P. zeylanica, Comminphora mukul, Allium sativum, Semecarpus anacardium, Hemidesmus indicus, Terminalia arjuna, Tinospora cordifolia. Withania somnifera and Ocimum sanctum, has antioxidant effects⁴⁸. It is known that in almost all cytotoxic effects of naphthoquinones, redox cycling is the most important process involved. In the presence of plumbagin, molecular oxygen can act as a univalent electron acceptor, generating superoxide, a reactive species that can damage various biomolecules. Antioxidant effects of aqueous/alcoholic extracts of P. zeylanica roots were studied to understand possible mechanisms of its action⁴⁷. Boiled ethanolic extracts and boiled aqueous extracts were most efficient. These extracts also significantly inhibited lipid peroxidation induced by cumene hydroperoxide, ascorbate-Fe²⁺ and peroxy-nitrite and contained high amounts of polyphenols and flavonoids. Protective effect of P. zeylanica was reported⁴⁹ against cyclophosphamide-induced genotoxicity and oxidative stress in Swiss albino mice

3.3 Antiulcer Activity

The anti-ulcer action of aqueous root extracts of *P. zeylanica* was studied on aspirin and indomethacin induced acute gastric ulceration in albino rats⁵⁰. The extract at doses, 25, 50 and 100 ml/kg observed statistically important (p < 0.05) dose dependent inhibition of aspirin induced gastric mucosal damage while in the indomethacin induced ulcer 50 and 100 mg/kg respectively proved statistically significant (p<0.05) inhibition. Oral acute toxicity testing showed oral LD₅₀ to be greater than 5000 mg/kg which revealed the wide margin of safety of root extracts of *P. zeylanica*⁵¹.

3.4 Anticancer Activity

P. zeylanica has been recommended in therapy of cancer in Siddha system of medicine⁵². Earlier work in Indian National Cancer Institute, Bethesda, Maryland, USA, has indicated that naphthoquinones from this plant are associated with anticancer activity⁵³. Plumbagin at I and IO µg /ml blocked mitosis in chick embryo fibroblasts in vitro⁵⁴. Plumbagin when administered intra tumour and orally at 2 mg/kg body weight brings about 70% and 60% relapse of tumor (fibrosarcoma) respectively¹⁶. Plumbagin is active for lymphocytic leukemia at 4 mg/kg body weight. Antitumor activity was also found against Dalton's ascitic lymphoma in mice by enhancing mean survival time and peritoneal cell counts⁵⁵. β -sitosterol from *P. zeylanica* showed cytotoxic activity on the human melanoma cell line (Bowes cells). Plumbagin β -sitosteryl-3 β -glucopyranoside-6'-O palmitate showed cytotoxic activity on both human cell lines MCF7 (Breast cancer cells) and bowes melanoma cells²⁶. Plumbagin suppressed growth of Raji (erythroleukemia), Calu-I (human lung carcinoma cell line), Hela (human cervical carcinoma cell line) and Wish (transformed epithelial cell line) tumor cell lines²³. Cytotoxic activity of bsitosteryl-3b- glucopyranoside-ad-O-palmitate from P. zeylanica was observed against MCF7 and Bowes cancer cell lines. b-Sitosterol inhibited Bowes cell growth and plumbagin was cytotoxic against MCF7 and Bowes cells²⁶. Plumbagin was found to be a potential novel agent in the control of hormone-refractory prostate cancer which is the second leading cause of cancer-related deaths in men⁵⁶. Plumbagin inhibits multiple molecular targets including PK Cepsilon, a predictive biomarker of Prostate cancer

aggressiveness⁵⁷. Stable plumbagin nanoparticles from P. zeylanica root extract were explored as a potential natural drug against prostate cancer. Inhibitory effect of the nanoparticles on the migration properties of prostate cancer cells revealed its therapeutic potentials for prostate cancer⁵⁸. Plumbagin can inhibit cell proliferation, block cell cycle, and induce apoptosis of APL cell line NB4 cells⁵⁹. Plumbagin is a powerful inhibitor of the NF-KB (nuclear factor kappa-light-chain-enhancer of activated B cells) activation pathway which leads to suppression of NF-KBregulated gene products, which explains its cell growth modulatory, anticarcinogenic, and radiosensitizing effects⁶⁰. Ethanolic extract of P. zeylanica possess substantial anticancer action against Ehrlich Ascites Carcinoma in animal models, and it decreases elevated level of lipid peroxidation having presence of higher terpenoids and flavonoids⁶¹. Plumbagin repressed the BAX, BCL-2, procaspase-3 expression, and cleaved caspase-3 in gastric cancer cells. Plumbagin inhibited the apoptosis in human gastric cancer cells due to its ability to suppress the STAT3 and Akt phosphorylation 62,63.

3.5 Leishmanicidal Activity

The quinones in *P. zeylanica* have promising antileishmanial activity against amastigotes of *Leishmania Donovani* and *L. amazonensis*. Plumbagin and its dimers, 3, 3'-bisplumbagin and 8, 8'-bisplumbagin have been used in the treatment of cutaneous leishmaniasis in *Amazonian Bolivia*⁶⁴.

3.6 Antifertility Activity

Extracts of Plumbago roots, when applied to ostium uteri, caused abortion^{65,66}. In albino rats plumbagin showed antiimplantation and abortifacient activities when given orally (1-2 mg/100gm body wt) without showing teratogenic effect. Application of plumbagin in doses 0.005-5µg, prevented oocyte development and affected fecundity and fertility in housefly Musca domestica67. Plumbagin demonstrated strong anti-progestational activity 68. Its root powder was 100% abortificient and showed 75% antiimplantation effects in rats. The antiimplantation effects depend on doses as well as initiation of treatment on specific days of pregnancy. Dose dependent antiimplantation response was 40-45% and 75%. The abortifacient response was 100% and dependent on mode of treatment in relation to the days of pregnancy⁶⁹. The plumbagin-free alcohol extract of root of P. zeylanica possesses antifertility activity in rats and is free from adverse actions^{70,65}. P. zeylanica therapy during the first 7 days of pregnancy abolished uterine proteins of 13,000, 19,000 and 26,000 and 75,000 Dalton molecular weights resulting in pre-implantationary loss. Proteins having molecular weights 55,000 and 65,000 Dalton were absent in aborted rats that were given P. zeylanica root powder from day 6 to day 17 of pregnancy⁷¹. Anti-implantation and abortifacient activity were reported in albino rats without any teratogenic effect of plumbagin in the doses of Img/100g⁷².

3.7 Antimalarial Activity

Plumbagin from *P. zeylanica* was reported to show potent antimalarial activity against *Plasmodium falciparum* both in mice and *in vitro* by regulating lipid peroxidation mechanism^{73,74}. The activity of *Plasmodium falciparum* enzyme, succinate dehydrogenase has been 50% inhibited by plumbagin at an inhibitory concentration of 5mM. It also prevented the *in vitro* growth of the parasite with a 50% inhibitory concentration of 0.27mM.

3.8 Antidiabetic Activity

Methanol extracts of *P. zeylanica* (root), had displayed mixed inhibition to alpha-glucosidase enzyme activity with 100% inhibition with the IC50 value of 3.46 µg/ml^{75.} Plumbagin (15 and 30 mg/ kg b wt) was orally administered to streptozotocin-induced diabetic rats for 28 days. An oral glucose tolerance test was performed on21st day. Plumbagin drastically lowered the blood glucose and substantially improved all other biochemical parameters to near normal. Plumbagin improved the activity of hexokinase and reduced the activities of glucose-6- phosphatase and fructose-1,6-bisphosphatase considerably in treated diabetic rats. Enhanced GLUT4 mRNA and protein expression were observed in diabetic rats after treatment with plumbagin⁷⁶.

3.9 Hypolipidaemic Activity

Ethanolic extract (50% v/v) of roots of *P. zeylanica* alone and combined with vitamin E, an antioxidant, in hyperlipidaemic rabbits, showed significant decrease in serum total cholesterol, LDL cholesterol and triglyceride levels. Plumbagin lowered serum cholesterol and LDL-cholesterol, by 53-86% and 61-91% respectively. It lowered cholesterol / phospholipid ratio by 45.8% and elevated the decreased HDL-cholesterol significantly⁷⁷. Marked reduction was seen with formulation of *P. zeylanica* and vitamin E. The total cholesterol/HDL and LDL/HDL cholesterol ratios were found significantly lowered. These results indicate that *P. zeylanica* extract contain hypolipidaemic and antioxidant substances and its use as a therapeutic tool in hyperlipidaemic subjects will be of benefit and promote further investigation in this field.

3.10 Trypanocidal Activity

Plumbagin from *P. zeylanica* exhibited high potency (IC 90= I- $5\mu g/ml$) against six strains of *Trypanosoma cruzi epimastigotes*, while the dimer 3, 3'-bisplumbagin and 8, 8'-bisplumbagin were less effective, with IC 90 in the 25-100 $\mu g/ml$ range⁶⁴.

3.11 Anti-inflammatory activity

P. zeylanica has various medicinal properties and is used in formulations of several ayurvedic compounds to treat inflammatory disorders such as rheumatoid arthritis and laryngitis⁷⁸. The phosphate buffered saline extract of roots of P. zeylanica was investigated for anti-inflammatory activity. The extract stabilized red blood cells subjected to heat. The extract exhibited a biphasic response. Enzymatic activities of both alkaline and acid phosphatases were reduced, while adenosine triphosphatase activity was stimulated in liver homogenates of formaldehyde induced arthritic rats⁷⁹. Analgesic and anti-inflammatory activity of hydroalcoholic extract of P. zeylanica leaf was reported^{80,81}. According to Chen and his co-workers extracts of P. zeylanica containing suberosin inhibited proliferation of human peripheral blood mononuclear cells through the modulation of the transcription factors NF-AT and NF-kappaB⁸² which provides

an explanation for the anti-inflammatory activity of *P. zeylanica*.

3.12 Anti Allergic Activity

The antiallergic properties of the 70% ethanol extract of *P. zeylanica* stems were studied. It inhibited systemic anaphylactic shock in mice, reduced homologous passive cutaneous anaphylaxis and skin reactions induced by histamine or serotonin in rats. Ethanol extract of *P. zeylanica* stems (50 μ g/ml) markedly increased intracellular cAMP content of rat mast cells. This extract inhibited mast cell-dependent immediate allergic reactions, mediated by reducing the release of mediators such as histamine from mast cells via elevating intracellular cAMP level and weakening the inflammatory action of mediators⁸³.

3.13 Anxiolytic Activity

The *in-vitro* anti-anxiety or anxiolytic activity of *P. zeylanica* in mice was associated with the anxiolytic drug diazepam. Activity of *P. zeylanica* leaf extracts was observed to be effective in mice. Male swiss albino mice between 8 - 10 weeks old weighing 20 - 25 gm were used in the research. A good response was observed in open field test⁸⁴.

4. ANTIMICROBIAL PROPERTIES OF PLUMBAGO ZEYLANICA

4.1 Antibacterial Activity

Extracts from roots of P. zeylanica showed antimicrobial properties. Aqueous extract and its partition (petroleum ether, dichloromethane, methanol, and aqueous residue) were effective against S. gallinarum, E. coli, P. vulgaris and K. pneumoniae⁸⁵. Aqueous and alcoholic extracts from roots of P. zeylanica demonstrated activity against B. subtilis, E. coli, P. vulgaris, S. typhimurium, P. aeruginosa and S. aureus. Among various medicinal plant extracts, alcoholic extract of P. zeylanica was found to show potentially interesting activity against pathogenic and opportunistic microorganisms⁸⁶. Alcoholic extract of P. zeylanica plant roots was tested against multidrug-resistant clinical isolates of bacteria, S. paratyphi, S. aureus, S. albus, E. coli, S. dysenteriae, K. pneumoniae and B. subtilis⁸⁷. The extract displayed strong antibacterial activity against all test bacteria irrespective of their antibiotic resistance behaviour. Phytochemical analysis of crude extract revealed the presence of flavonoids, saponins and naphthoquinones. Plumbagin along with some related naphthoquinones was found effective against E. coli, C. jejuni, Bacillus sp., Staphylococcus sp., Mycobacterium sp., C. diptheriae⁸⁸(Table 1). It has shown antibacterial activity against Acineobacter⁸⁹ Antibacterial activity of Plumbago root extracts reported against S. marcescence and P. mirabilis was reported for first time⁴². Anti-Helicobacter pylori activity of P. zeylanica was detected by ⁹⁰. Methanolic extract of P. zeylanica roots showed anti-bacterial effect against Bacillus subtilis ⁹¹. Ethyl acetate extract exhibited lowest minimum inhibitory concentrations against five H. pylori strains, of which ranged from 0.32 to 1.28 mg ml/l. Ethanolic extract of P. zeylanica showed anti-microbial activity against Salmonella typhi, Pseudomonas aeruginosa, Bacillus subtilis and Staphylococcus aureus ⁹².

4.2 Antifungal Activity

Broad spectrum antifungal activity of *Plumbago* root extracts was reported. Antifungal activity of *P. zeylanica* against *F. oxysporium* and *C. humicolus*, was reported for the first time⁴². *P. zeylanica* also showed antifungal activity against *P. notatum*, *P. canadense*, *R. nigricans*, and *E. floccosum* at concentration $10\mu g/ml^{16}$. High potency was observed in the extracts of *P. zeylanica* (4mg/ml) against *Candida* which indicates that the plant has a potential source for anticandidal drugs⁹³. *P. zeylanica* naphthoquinones were effective against *A. flavus*, a fungus that contaminates commercial products walnuts⁹⁴. The quinines delayed germination of fungus, its growth and aflatoxigenesis. A very dilute solution (i.e. a concentration of

1:50,000) of plumbagin was lethal to a wide spectrum of bacteria and pathogenic fungi, i.e. Coccidioides imminites, Histoplasma capsulatum, Trichophyton spp., C. albicans, A. niger and A. flavus⁹⁵.

4.3 Antiviral Activity

80% methanolic extracts of *P. zeylanica* exhibited antiviral activities against coxsackievirus B3 (CVB3), influenza A virus and herpes simplex virus typeI Kupka (HSV-I) using cytopathic effect (CPE) inhibitory assays in HeLa, MDCK, and GMK cells, respectively. It was confirmed with plaque reduction assays⁹⁶.

Table 1. Antimicrobial activity of plumbagin			
Organism	MIC of Plumbagin from P. zeylanica (µg/ml)		
Gram positive bacteria 16,97,98			
Bacillus subtilis	0.2		
Staphylococcus aureus	20		
S. citreus	20		
S. albus	20		
Gram negative bacteria ^{16, 97,99}			
Salmonella dublin	20		
Salmonella. paratyphi	20		
Klebsiella Pneumonia	20		
Pseudomonas aeruginosa	12.5		
Escherichia coli	50		
Rhinotricum nigricans	10		
Penicillium canadense	10		
Penicillium notatum	10		
Penicillium lilacinum	10		
Metarhizium nana	10		
Candida albicans	0.78		
Protozoa ⁴⁶			
Leishmania donovani	10 – 20		
Leishmania mexicana	10 – 20		

5. INSECTICIDAL AND LARVICIDAL ACTIVITY

Insects can act as vectors of various diseases. The control of insects is of great importance; mainly in developing countries where they are commonly endemic, most of them are transmitted zoonotically. Plumbagin affects insect growth, metamorphosis, lowers the ability of mating in males and has larvicidal activity (Table 2). Hexane and chloroform and hexane crude extracts of *P. zeylanica* showed highest larvicidal activity against *A. gambiae* i.e. LC50 6.4 and 6.7 μ g/ml respectively ¹⁰⁰. *P. zeylanica* extract possesses larvicidal activity against second, third, and fourth instar larvae of *Aedes aegypti*. LC (50) values of all the extracts in different solvents of *P. zeylanica* were less than 50 ppm against all tested larval¹⁰¹.

	Table 2. Effect of plumbagin on different insects			
Sr.	Insect	Activity		
No.				
1	Musca domestica	Affect insect growth and		
	(Diptera -Muscidae)	Metamorphosis ¹⁰²		
2	Dysdercus koenigii (Heteroptera - Pyrrhocoridae)	Lower growth rate and rise the time taken for molting. Reduce ability of mating in males and affect fecundity of females in freshly moulted adults ¹⁰³		
3	Dactylotum corallinum (Orthoptera -	Insect feeding deterrent ¹⁰⁴		

	Acrididae)	
4	Helicoverpa armigera (Lepidoptera - Noctuidae)	Insect growth regulator. Affect number of major protein bands in protein profiles of cuticle of treated larvae. It also affects neurosecretory cells ¹⁰⁵
5	Dysdercus koenigii (Heteroptera - Pyrrhocoridae)	Growth regulator ¹⁰⁶
6	Arachnis aulaea (Lepidoptera - Arctiinae)	Responsible for selectivity feeding Behavior ¹⁰⁴
7	Culex fatigans (Diptera - Culicidae)	Larvicidal activity ¹⁰⁷
8	Phoetaliotes nebrascencis (Orthoptera - Acrididae)	Insect feeding deterrent ¹⁰⁴
9	Sphenarium purpurascens (Orthoptera - Acrididae)	Insect feeding deterrent ¹⁰⁴
10	Culex quinquefasciatus (Diptera - Culicidae)	Larvicidal activity ¹⁰⁸

6. ANTIMUTAGENIC ACTIVITY

Antimutagenic activity of plumbagin from *P. zeylanica* was tested against known chemical mutagens in a standard mutagenicity test system of Ames using *S. typhimurium* strains¹⁰⁹. Plumbagin did not show any mutagenic effect, whereas it reduced significantly mutagenic effect of 4-nitrophenylene diamine, phenyl hydrazine and sodium azide in test strains of *S. typhimurium*, suggesting that plumbagin possessed antimutagenic activity. Actively growing *E. coli* cells when exposed to plumbagin, a redox cycling quinone which increases flux of O_2 radicals in the cell, were mutagenized by this treatment¹¹⁰. *E. coli* showed an inducible DNA repair response specific for the type of oxidative damage generated during incubation with plumbagin. Methanolic extracts of *P. zeylanica* roots exhibited varying levels of antimutagenicity¹¹¹.

7. FORMULATIONS AND PHARMACOLOGICAL ACTIVITIES OF PLUMBAGO ZEYLANICA:

The root extracts of Plumbago species have also been

incorporated in various Indian indigenous ayurvedic drug formulations, namely Chitrakadi vati, Chitraka-haritaki, Yakritaplihari Drakshasava, Dashamoolarishta, lauha, Ashwagandharishtha, Chitrakadi lauha, Lauhasava, Chitrakadi ghrita, Chitrakadi taila, Chitrakadi Lepa as well as Asokarishtam, Livosprin, Livomyn, Livokin etc¹¹² (Table 3). Importance of P. zeylanica and its possible pharmaceutical activity for the development of new herbal formulations had been evaluated¹¹³. P. zeylanica showed antipyretic, antibacterial, antifungal, antifertility, anticancer, anticoagulant, antitumor, hepatoprotective and cytotoxic activities¹¹⁴. Plumbagin given orally at 2 mg/kg, decreased tumor growth by 70%. Tropical application of plumbagin has been found to be useful in patients with common wart. Plumbagin¹¹⁵ has various pharmacological activities like antimalarial, cardiotonic etc¹¹⁶. It has been described in literature and is shown to possess a wide variety of bioactivities¹⁶. It shows activity against several grampositive bacteria, gram-negative bacteria as well as Candida species.

Table 3. Formulations with a root powder of Chitraka manufactured in India					
Name of formulation	Ingredients	% of P. zeylanica (Chitraka)	Use	Manufacturer	Reference
Chitrakadi vati	Chitraka, pimpali mool, Lavanani, Ajamodra, Hingu, Vyosham, Doxari, Chavya	12.5	Increases digestive capacity	Bharadwaj Pharmaceuticals, Baidyanath, Ayurved Rasashala, Unjha Pharmaceuticals, Kahdiwale Vaidya.	112
Chitraka-haritaki	Chitraka, Dashamoolakvath, Awala swaras, Guduchi swaras, Haritaki, Vyosh, Trijat,	20	Tuberculosis, cough, cold, worms, tumer, breathlessness	AVP (Kerala), as per demand	117

	Yavakshara, Shahad				
Dashamoolarishta	Dashamool, Danti, Chitraka mool, Haritaki	20	Piles, colitis, anemia, to increase digestive capacity	Arkashala, Sandu Brothers, Bharadwaj Pharmaceuticals, Baidyanath, Ayurved Rasashala, Unjha Pharmaceuticals, Kahdiwale Vaidya.	112
Yakritaplihari lauha	Hinguloth parad, Gandhak, Loha, Abrhakam, Tamra bhasma, Manashila, Rajani choorna, Jaipal, Takan, Sheelajatu, Danti, Chitraka, Nirgundi, Adrak, Brungaraj ras.	2 - 5	All types of ascitis. Increases digestive capacity.	As per demand	117
Drakshasava	Draksha, Madha, Khadisakhar, Dhataki, Kankol, Lavang, Jaiphal, Mire, Pimpli, Chitraka, Chavya, Pimpal mool, Renuka	5	Worms, skin diseases, tonic, wounds, eye infections etc.	Arkashala, Sandu Brothers, Bharadwaj Pharmaceuticals, Baidyanath, Ayurved Rasashala, Unjha Pharmaceuticals, Kahdiwale Vaidya.	112
Lauhasava	Lohachurna, Trikatu, Triphala, Yavani, Widang, Mustaka, Chitraka, Dhatki Phool, Shahad, Gud, Shudhajal	Ι	Ascitis, splenic disorders, itching, cardiac disorders	Arkashala, Sandu Brothers, Bharadwaj Pharmaceuticals, Baidyanath, Ayurved Rasashala, Unjha Pharmaceuticals, Kahdiwale Vaidya.	111
Ashwagandharishtha	Ashwagandha, Shwetamusali, Manjishtha, Harithaki, Rajani, Daru halad, Madhuk, Rasna, Vidari, Arjun, Trivruta, Chandana shwet, Chandana rakta, Wacha, Chitraka, Ananta, Nishottar, Dhatki, Madha, Trikatu, Trijat, Nagkeshar	I	Coma, epilepsy, weakness	Arkashala, Sandu Brothers, Bharadwaj Pharmaceuticals, Baidyanath, Ayurved Rasashala, Unjha Pharmaceuticals, Kahdiwale Vaidya.	112
Chitrakadi lauha	Chitraka, Nagar, Vasa, Shalparni, Talpushpa, Apamarg, Manak, Loha bhasma, Abhraka Bhasma, Pimpli Choorna, Tamra Bhasma, Javakahara, Sendhav, Sarvalavana, Gomutra	20	Splenomegaly, hepatic disorders, fever, jaundice, edema, dysentery	As per demand	112
Chitrakadi ghrita	Chitraka, Takra, Kshira	30	Dysentery, digestive disorders	AVP (Kerala), Baidyanath	112
Chitrakadi taila	Chitraka arka, Trivruta, Patha, Bakuchi, Ashwamar, Sudha, Wacha, Kalihari, Saptaparni, Sajjikshar, Jyotishmati	9	Fistula, wound healing, skin disorders	AVP (Kerala)	112
Chitrakadi Lepa	Chitraka, Ela, Bimbi, Wasa, Arka, Suntha	16	Rash type skin diseases	As per demand	112

8. ACTION OF PLUMBAGIN

Charak has described Chitraka as Deepan-pachan dravya (useful in digestion) $^{\rm 117}.$ In fact, practitioners of Indian system

of medicine have been using drugs in the form of a decoction or powder for centuries. A chemical substance 'Plumbagin' was first isolated in 1829 by¹¹⁸ Dulong D Astafort form *P. zeylanica* and first synthesized by Fieser and

Dunn after a century in 1936¹¹⁹. Its active principle content is an alkaloid "Plumbagin" which stimulates secretion of sweat, urine and bile and has stimulant action on the nervous system. Plumbagin is quinine and is capable of abstracting electrons from electron transfer components and diverts the electron flow to molecular dioxygen to form superoxide radicals¹²⁰. 0_2 ⁻ Gives rise to OH radicals and H_2O_2 by enzymatic and non-enzymatic reactions responsible for damage to micromolecules including DNA in microorganisms. Non-DNA damaging concentrations of plumbagin diminished the DNA damage induced by catechol, which provides support for the idea that plumbagin may act as an antioxidative agent at low concentrations¹²¹.Plumbagin induced mammalian topoisomerase II-mediated DNA cleavage in vitro⁹³. Treatment of a reaction mixture containing this naphthoquinone and topoisomerase II at an elevated temperature of 65 $^{\circ}$ C resulted in a great reduction in DNA cleavage. Plumbagin has anticancer, antileishmanial, antibacterial, antifungal properties as well as a contraceptive effect. It has a potential as a chemotherapeutic agent⁵⁶. It also has cardiotonic action¹¹⁶, insecticidal activity¹²², hypolipidemic and anti-atherosclerotic effect by reducing the level of cholesterol and LDL cholesterol in rabbit¹²³. Plumbagin augments macrophage bactericidal activity by potentiating oxyradical release at low concentration whereas at higher concentration it has inhibitory activity¹²⁴. Plumbagin when administered orally, at a dosage of 4 mg/kg body weight induces tumor regression in 3-methyl-4dimethyl amino azobenzene (3MeDAB) induced hepatoma in Wistar male rats⁵⁶. Certain gluconeogenic enzymes, glucose-6-phosphatase and fructose namely, -1.6diphosphatase decreased in tumor hosts, whereas plumbagin administration increased gluconeogenic enzyme levels in treated animals. These investigations indicate the molecular basis of different biological behavior of 3MeDAB induced hepatoma and anti-carcinogenic property of plumbagin against hepatoma studied in rats. When tested against the resistant strain of M. tuberculosis H37Rv, plumbagin exhibited inhibitory activity at <12.5 microg/mL¹²⁵. The major effects of plumbagin on chick embryo fibroblast cultures were arrest of cell growth and proliferation decrease in mitotic index with accumulation of cells in metaphase at I µg concentration. There was indication of chromosomal aberrations like clumping of chromosomes, with degeneration as shown by nuclear and cytoplasmic vacuolization and nuclear polymorphism. Plumbagin at lower concentration behaves like a spindle poison by preventing entry of cells into mitosis like colchicines but at higher concentrations it exhibits nucleotoxic and cytotoxic effects. Plumbagin effects on reproductive function of rat. It causes selective testicular lesions in dogs. The wet weight of testes and epididymides decreases. Plumbagin inhibits spermatogenesis. It results in significant decrease in protein and RNA contents of testes and epididymides associated with loss in weight of these organs⁵⁴.

9. PLASMID CURING BY P. ZEYLANICA ROOT EXTRACT, PLUMBAGIN AND LAWSONE

Development and spread of antibiotic resistance are problems with prolonged chemotherapy against bacterial infections. Elimination of plasmid mediated drug resistance in pathogenic strains of bacteria is of great importance both, in treatment of bacterial infection and in microbial genetics. The already ineffective therapy can be made effective by converting resistant cells into sensitive ones¹²⁶. Reversal of drug resistance by plumbagin has been recorded in microorganisms¹²⁷. DNA strand session and plasmid curing activity of an Indian folk medicine constituent Chitrak has been previously reported¹²⁸. P. zeylanica extract cured plasmid from 14% E. coli (pUK 651) treated cells, confirmed by determining the loss of resistance markers in cured derivative culture ⁸⁷. The root extracts cured plasmid encoded antibiotic resistances from the clinical isolates and reference strains at curing efficiencies of 4 to 100%. Petroleum ether root extract of P. zeylanica demonstrated higher plasmid curing activity and was higher than known plasmid curing agents like ethidium bromide or acridine orange¹²⁹. Plumbagin was effective in selectively eliminating stringent, conjugative, multidrug-resistant plasmids from E. coli strains. Simultaneous loss of resistance to antibiotics in plumbagin-treated cell indicated loss of plasmid⁹⁹. Plumbagin at 50µg/ml cured silver resistance from Acinetobacter baumannii BL88 at a frequency of 69%. Along with Agr other markers i.e Cdr, Sbr, Apr and Smr were also cured at varying frequencies¹³⁰. Plumbagin was found to be effective in curing the plasmids pUPI 102(Tc r, Cm r, Hg r) and pUPI 103(Pn r, Cb r, Km r) from antibiotic and metal resistant strains of Acinetobacter baumannii ¹³¹. Plumbagin at subinhibitory concentration 62.5 µg/ml eliminated the plasmid pUPI200(Sm r, Km r, Gm r) from E. coli K12 [53.2 with 21% curing efficiency¹³². A. baumannii C11, a soil isolate exhibited high level of resistance to multiple antibiotics and heavy metals. Plumbagin eliminated resistances at following efficiencies: Gmr (100%), Nm (100%), Cnr (80%), Ctr (100%), Cmr (74%), Tpr (100%), Pnr (18%), Agr (100%), Hgr (100%), Cdr (100%)¹³³. Plasmid pUPI102 (Gmr, Nmr, Tcr, Hgr) from A. junii was cured by plumbagin¹³⁴. Plumbagin was found to ACN4 be far more effective as a plasmid curing and antibacterial agent than its metal complexes. It caused DNA strand scission¹²⁸. Plumbagin has been used in plasmid elimination from antibiotic resistant clinical strains of Acinetobacter 135. Plasmids pUPI275 (Smr, Sdr) and pUPI276 (Agr, Apr, Cbr, Tcr, Cmr,) from A. baumannii BL54 were cured by treatment with plumbagin^{136,137}. Plumbagin intercalates into DNA molecule and induces topoisomerase-II mediated DNA cleavage in vitro97. Curing ability of plant extract is due to plumbagin as it intercalates with DNA molecule and inhibits plasmid replication selectively at sub-MIC concentration. It is dependent on ability of plumbagin to undergo redox cycling to produce superoxide that can damage various macromolecules (Table 4). In a research investigation,²⁸ Patwardhan and her coworkers purified a compound Lawsone from P. zeylanica roots able to eliminate antibiotic resistance and cure plasmids from pathogenic strains resistant to multiple antibiotics without any ill effect on mammalian cells. The synergistic effect of lawsone with the antibiotic exhibited its tremendous potential in modern day therapeutics. The non-toxic, non-mutagenic, plasmid curing and plasmid transfer inhibiting role of lawsone made it a potential drug of choice in the treatment of antibiotic resistant bacterial strains, demonstrating a new dimension in antibiotic therapy.

Table 4. Plasmid curing by P. zeylanica root extract, Plumbagin and Lawsone in Acinetobacter and E. coli				
Strain	Plasmid	Resistance markers cured	Reference	
A. baumannii CII	_P UPI 101	Cd ^r	89	
A. junii ACN4	_P UPI 102	Gm ^r , Nm ^r , Tc ^r , Hg ^r	89	

A. baumannii CAII4	_P UPI 104	Ap ^r , Km ^r	89
A. baumannii APH5	PUPI 105	Cd ^r	89
A. baumannii APH5	pUPI 106	Pn ^r	89
A. baumannii APH5	PUPI 107	Cb ^r	89
A. baumannii B32	_P UPI 108	Pn ^r , Ap ^r , Cb ^r , Am ^r , Cp ^r , Cm ^r , Km ^r	89
A. baumannii B32	_P UPI 109	Tc ^r , Sm ^r , Hg ^r ,	89
A. baumannii B32	_P UPI I I 0	Cu ^r ,	89
A. baumannii CII	_P UPI I I I	Pn ^r , Cb ^r , Km ^r	89
A. baumannii CII	PUPI 112	Cp ^r , Nm ^r , Hg ^r	89
A. baumannii BLII0	_P UPI 200	Sm ^r , Km ^r , Gm ^r , Cp ^r	89
A. baumannii CAII4	R751	Tp ^r , Su ^r	89
A. baumannii BL88	_P UPI 199	Ag ^r , Cd ^r , Sb ^r , Ap ^r , Sm ^r	136
A. baumannii BL54	_P UPI 275	Sm ^r , Cd ^r	136
A. baumannii BL54	_P UPI 276	Ap ^r , Tc ^r , Cm ^r , Ag ^r	136
A. baumannii A24	pUPI281	St ^r , Ap ^r , Gm ^r , Ak ^r	28
E. coli 46R641	Tp181	Ap ^r , Cm ^r , Km ^r , Sm ^r , Su ^r , Tc ^r	99
E. coli 48R371	R162	Ap ^r , Cm ^r , Sm ^r , Su ^r , Tc ^r	99
E. coli 44R266	R6K	Ap ^r , Sm ^r ,	99
E. coli 42R873	TP154	Ap ^r , Km ^r , Tc ^r	99
E. coli HB101	pBR322	Ap ^r , Tc ^r	99
E. coli HB101	pBR329	Ap ^r , Cm ^r , Tc ^r	99
E. coli 391	RP4	Ap ^r , Km ^r , Tc ^r	138
E. coli 393	_Р КТ231	Km ^r , Sm ^r	138
E. coli 398	pRK2013	Ap ^r , Km ^r ,	138
E. coli	PUK651	Ap ^r , Km ^r , Co ^r	87
E. coli	PRK2013	Ap ^r , Km ^r	28

Ap-Ampicillin; Tc-Tetracycline; Km-Kanamycin; Sm-Streptomycin; Cb-Carbenicillin; Pn-Penicillin; Su-Sulphonamide; Cm-Chloramphenicol; Nm-neomycin; Tp-Trimethoprim; Cp-Ciprofloxacin; Gm-Gentamycin; Co-Cobalt; Hg: Mercury

10. DISCUSSION

From primordial times of Vedas, Charaksamhita and Sushrutsamhita, to present day advances and research in medicine, the therapeutic importance of Chitraka i.e., Plumbago zeylanica as an excellent Indian remedy, have been upheld through test of time. Therefore, now it is very much essential to make further efforts to explore about nature and utilize P. zeylanica plant for betterment of mankind in today's age of infections and pollutions. In fact, today there is a need to combine branches of allopathic medicine with ayurvedic science for benefit of mankind. Formulations and preparations of P. zeylanica roots, their effects and pharmacological activities were studied. One of the ways to overcome antibiotic resistance problem is to eliminate genes encoding antibiotic resistance in bacteria. Because of toxicity of other curing agents like acridine orange and ethidium bromide, there is a constant need of developing novel curing agents which are more effective and at the same time nontoxic. Lawsone and plumbagin could eliminate antibiotic resistance and cure plasmids from pathogenic strains that are resistant to multiple antibiotics without any ill effect on mammalian cells at lower concentration. Goal of this review was to explore the significance of P. zeylanica and its potential medicinal and plasmid curing activity for the development of herbal formulations. The findings and outcomes of new this research would be useful in using plumbagin and lawsone from P. zeylanica roots as potential drugs of choice in the treatment of antibiotic resistant bacterial strains. These findings are of particular significance as plasmid encoded antibiotic resistance is a major challenge for physicians to treat. Already ineffective antibiotics could

become effective if plasmid encoded antibiotic resistance is eliminated from the population. *P. zeylanica* root extracts can eliminate the plasmid encoded antibiotic resistance and render the cell sensitive to the antibiotics. The ineffective or outdated antibiotics could be rejuvenated if used in combination with such curing agents like plumbagin and lawsone. This would be a novel approach towards controlling multidrug resistant bacterial infections especially in hospital environment. Clarification of exact mechanism by which lawsone triggered plasmid curing in bacterial cells is not known at present and requires further extensive investigation.

II. CONCLUSION

P. zeylanica is used from centuries in Ayurvedic medicine. It is a valuable medicinal plant universally used in herbal formulations. It is chemically rich with its diverse contents including many active secondary metabolites like plumbagin and lawsone. The pharmacological attributes of P. zeylanica have been revalidated in modern-day sciences through several in vivo and in vitro studies. The present investigation elaborates broad spectrum medicinal properties of Plumbago zeylanica including antioxidant, antiulcer, anticancer, leishmanicidal, antifertility, antimalarial, antidiabetic, hypolipidaemic, trypanocidal, antibacterial, antifungal, antiviral, anti-inflammatory, antimutagenic, antiallergic, larvicidal, insecticidal, wound healing and anxiolytic activities. This plant has immense potential as a plasmid curing agent. The present study reveals applications of root extracts of P. zeylanica as plasmid curing agents to contain the infections and the spread of antibiotic resistance especially in hospital environment. Plasmid elimination

activity of *P. zeylanica* root extracts, plumbagin and lawsone has been documented for the first time in the present review.

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14. REFERENCES

- Cowan MM. Plant products as antimicrobial agents. Clin Microbiol Rev. 1999;12(4):564-82. doi: 10.1128/CMR.12.4.564, PMID 10515903.
- Edeoga HO, Okwu DE, Mbaebie BO. Phytochemical constituents of some Nigerian medicinal plants. Afr J Biotechnol. 2005;4(7):685-8. doi: 10.5897/AJB2005.000-3127.
- Santos PRV, Oliveira ACX, Tomassini TCB. Controle microbiógico de produtos fitoterápicos. Rev Farm Bioquím. 1995; 31:35-8.
- 4. Sandhu DS, Heinrich M. The use of health foods, spices and other botanicals in the Sikh community in London. Phytother Res. 2005;19(7):633-42. doi: 10.1002/ptr.1714, PMID 16161027.
- Eloff JN. Which extractant should be used for the screening and isolation of antimicrobial components from plants? J Ethnopharmacol. 1998;60(1):1-8. doi: 10.1016/s0378-8741(97)00123-2, PMID 9533426.
- 6. Neu HC. The crisis in antibiotic resistance. Science. 1992;257(5073):1064-73.
 - doi: 10.1126/science.257.5073.1064. PMID 1509257.
- Mulligan ME, Murray-Leisure KA, Ribner BS, Standiford HC, John JF, Korvick JA, Kauffman CA, Yu VL. Methicillin-resistant Staphylococcus aureus: a consensus review of the microbiology, pathogenesis, and epidemiology with implications for prevention and management. Am J Med. 1993;94(3):313-28.
 - doi: 10.1016/0002-9343(93)90063-u, PMID 8452155.
- 8. Ahmad I, Yadava JNS, Ahmad S. High level transfer among E. coli. Indian J Anim Sci. 1994;4:439-45.
- Huebner RE, Wasas A, Mushi A, Mazhani L, Klugman K. Nasopharyngeal carriage and antimicrobial resistance in isolates of Streptococcus pneumoniae and Haemophilus influenzae type b in children under 5 years of age in Botswana. Int J Infect Dis. 1998;3(1):18-25.
 - doi: 10.1016/S1201-9712(98)90090-X.
- Gogate VM. Pune: Dravyagunavidnyan, Continental Publication; 1982. p. 302.
- Ray P, Gupta HN. 'Charaka Samhita'. A Scientific synopsis. New Delhi: Indian National Science Academy; 1980.
- Ray P, Gupta HN, Ray M. 'Sushruta Samhita'. A Scientific synopsis. New Delhi: Indian National Science Academy; 1980.
- Chopra RN, Chopra IC, Handa KL, Kapur LD. Chopra's indigenous drugs of India. 2nd ed. Calcutta: UNITED nations Dhur & Sons; 1958. p. 12-6.
- Chopra RN, Nayer SL, Chopra IC. Glossary of Indian medicinal plants. 3rd ed. New Delhi, India: Council of Scientific and Industrial Research; 1992. p. 197-8.
- 15. Singh S, Priyadarshi A, Chaudhary SP, Singh B, Sharma P. Pharmacognostical and phytochemical evaluation of

Ayurved, Bharati Vidyapeeth University, for provision of valuable references from ayurvedic science.

13. CONFLICT OF INTEREST

Conflict of interest declared none.

Chitraka (Plumbago zeylanica Linn.). J Pharm Innov. 2018;7(6):281-5.

- Krishnaswamy M, Purushothaman KK. Plumbagin: A study of its anticancer, antibacterial, and antifungal properties. Indian J Exp Biol. 1980;18(8):876-7. PMID 7461745.
- Thakur RS, Puri HS, Husain A. Major medicinal plants of India. Lucknow, India: Central Institute of Medicinal and Aromatic Plants; 1989.
- Chauhan M. A review on Morphology, Phytochemistry and Pharmacological activities of medicinal herb Plumbago zeylanica Linn. J Pharmacogn Phytochem. 2014;3(2):95-118.
- Vander-Vijver LM. (). Distribution of plumbagin in the mplumbaginaceae. PhytoChemistry. 1972;11(11):3247-8.

doi: 10.1016/S0031-9422(00)86380-3.

- Dinda B, Hajara AK, Chel G. Naphthoquinones of Plumbago species: a review. J Indian Chem Soc. 1997; 74:974-9. J-Global ID: 200902148576517075.
- Tyagi R, Menghani E. A review on Plumabgo zeylanica: A compelling herb. Int J Pharm Sci Res. 2014;5(4):119-26.
- 22. Yuvaraj D. Mandavkar YD, Jalalpure SS. A comprehensive review on Plumbago zeylanica Linn. Afr J Pharm Pharmacol. 2011;5(25):2738-47. doi: 10.5897/AJPP11.739.
- Lin LC, Yang LL, Chou CJ. Cytotoxic naphthoquinones and plumbagic acid glucosides from Plumbago zeylanica. PhytoChemistry. 2003;62(4):619-22. doi: 10.1016/S0031-9422(02)00519-8.
- Qian XL, Xiao TC, Pu-Zhu HH. Study on the chemical constituents of Plumbago zeylanica Linn. I. Isolation and identification of the constituents of Plumbago zeylanica Linn. Hua Hsveh Hsueh Pao.1980;38(4):377-80.
- 25. Dinda B, Saha S. Chemical constituents of P. zeylanica and Thevetia neriifolia roots. J Indian Chem Soc. 1990;67:88-9.
- Nguyen AT, Malonne H, Duez P, Vanhaelen-Fastre R, Vanhaelen M, Fontaine J. Cytotoxic constituents from Plumbago zeylanica. Fitoterapia. 2004;75(5):500-4. doi: 10.1016/j.fitote.2004.03.009.
- 27. Zhang QR, Mei ZN, et al. Chemical constituents from aerial parts of Plumbago zeylanica Linn. Zhong Yao Cai. 2007;30(5):558-60.
- Patwardhan RB, Dhakephalkar PK, Chopade BA, Dhavale DD, Bhonde RR. Purification and characterization of an active principle, lawsone, responsible for the plasmid curing activity of Plumbago zeylanica Root Extracts. Front Microbiol. 2018;9:2618. doi: 10.3389/fmicb.2018.02618, PMID 30467495.

- Anonymous. Medicinal plants of India. Vol. II. New Delhi: Indian Council of Medical Research; 1987p. p. 471-7.
- 30. Dinda B, Saha S. The free amino acids of Plumbago zeylanica. J Indian Chem Soc. 1987;64(4):261.
- Samhita C. Chikitsasthan. Adhyaya. 6th ed Tripathi B, Chaukhambha Vidyabhavan V, editors. Vol. 15; 1999. p. 102-3.
- 32. Li HL. Flora of Taiwan. 2nd ed. Editorial Committee of the flora of Taiwan; 1998. p. 79.
- Patwardhan RB, Dhakephalkar PK, Chopade BA. Medicinal importance of Chitraka plant: a review. Srushti health bulletin. 2005; 4:4-5.
- Sharma PC, Yelne MB, Dennis TJ. Plumbago zeylanica in Database on medicinal plants used in Ayurveda (Central Council for Research in Ayurveda and Siddha. New Delhi; 2000. p. 102-13.
- 35. Anonymous. Study of Plumbago zeylanica, materia medica, vegetable section, Murugesa Mudaliar. Government of Tamil Nadu; 1969p. p. 311-3.
- 36. Anonymous. The ayurvedic pharmacopoeia of India, Ministry of Health & Family Welfare, Govt, of India, New Delhi. Part j. 1989p;I:29.
- 37. Rajakrishnan R, Lekshmi R, Benil PB, Thomas J, AlFarhan AH, Rakesh V, Khalaf S. Phytochemical evaluation of roots of Plumbago zeylanica L. and assessment of its potential as a nephroprotective agent. Saudi J Biol Sci. 2017;24(4):760-6. doi: 10.1016/j.sibs.2017.01.001.
- Nadkarni KM. Plumbago zeylanica. Indian Materia Medica. Mumbai: Popular Press Prakashan Pvt Ltd.. 1976; I: 990.
- Sepúlveda-Boza S, Cassels BK. Plant metabolites active against Trypanosoma cruzi. Planta Med. 1996;62(2):98-105.

doi: 10.1055/s-2006-957827.

- Chan-Bacab MJ, Peña-Rodríguez LM. Plant natural products with leishmanicidal activity. Nat Prod Rep. 2001;18(6):674-88. doi: 10.1039/b100455g. Durga R, Shridhar P, Polasa H. Effects of plumbagin on antibiotic resistance in bacteria. Indian J Med Res. 1990; 91:18-20.
- Patwardhan RB. Herbal naphthoquinones as antimicrobial and plasmid curing agents. Available from: http://lib.unipune.ac.in:8080/xmlui/bitstream/handle/12 3456789/1508/01_ [PhD thesis] Submitted to University of Pune: India; 2007.
- 42. Devarshi P, Patil S, Kanase A. Uterine protein patterns in Plumbago zeylanica root powder and 17 beta estradiol treated albino rats. Indian J Comp Anim Physiol. 1990;8(1):1-7.
- 43. Devarshi P, Patil S, Kanase A. Effect of Plumbago zeylanica root powder induced preimplantationary loss and abortion on uterine luminal proteins in albino rats. Indian J Exp Biol. 1991;29(6):521-2. PMID 1889824.
- Gupta V, Yadav SK, Singh D, Gupta N. Evaluation of wound healing activity of herbal drug combination of Rubia cordifolia, Centella asiatica, Terminalia belerica, Plumbago zeylanica and Withania somnifera. Int J Pharm Life Sci. 2011;2(7):952-4.
- 45. Kodati DR, Burra S, Kumar GP. Evaluation of wound healing activity of methanolic root extract of

Plumbago zeylanica L. in Wistar albino rats. Asian J Plant Sci Res. 2011;1(2):26-34.

- Tilak JC, Adhikari S, Devasagayam TPA. Antioxidant properties of Plumbago zeylanica, an Indian medicinal plant and its active ingredient, plumbagin. Redox Rep. 2004;9(4):219-27. doi: 10.1179/135100004225005976.
- 47. Mary NK, Babu BH, Padikkala J. Antiatherogenic effect of Caps HT2, a herbal Ayurvedic medicine formulation. Phytomedicine. 2003;10(6-7):474-82. doi: 10.1078/094471103322331412.
- Sivakumar V, Niranjali Devaraj S. Protective effect of Plumbago zeylanica against cyclophosphamide-induced genotoxicity and oxidative stress in Swiss albino mice. Drug Chem Toxicol. 2006;29(3):279-88. doi: 10.1080/01480540600652921.
- Falang KD, Uguru MO, Wannang NN, Azi IH, Chiamaka N. Anti-ulcer activity of Plumbago zeylanica Linn root extract. J Nut Prod Plant Resour. 2012;2(5):563-7.
- Sharma A, Singh N. A multifarious potent herb: Plumbago zeylanica: A mini review. Int J Recent Sci Res. 2015;6(6):4825-9.
- 51. Mudaliar M. Materia medica. India: government of Tamil Nadu; 1969. p. 311-4.
- Carter SK, Newman JW, Rall D, Schein P, Cooney DA, Davignan JP, Davis RD, Murray BR, Schepartz SA, Vanditti JM. NIH document NSC.11905: 1967; p 1-13.
- Santhakumari G, Saralamma PG, Radhakrishnan N. Effect of plumbagin on cell growth and mitosis. Indian J Exp Biol. 1980;18(3):215-8. PMID 7390548.
- Kavimani S, Lyengar R, Madheswaran M, Jayakar B, Gupta M, Majumdar UK. Antitumor activity of plumbagin against Dalton's ascitic lymphoma. Indian J Pharm Sci. 1996;58(5):194-6.
- Parimala R, Sachdanandam P. Effect of plumbagin on some glucose metabolising enzymes studied in rats in experimental hepatoma. Mol Cell Biochem. 1993;125(1):59-63. doi: 10.1007/BF00926835.
- 56. Aziz MH, Dreckschmidt NE, Verma AK. Plumbagin, a medicinal plant-derived naphthoquinone, is a novel inhibitor of the growth and invasion of hormone-refractory prostate cancer. Cancer Res.

2008;68(21):9024-32. doi: 10.1158/0008-5472.CAN-08-2494.

57. Kapare HS, Metkar SR, Shirolkar SV. Anticancer potential of Plumbago zeylanica Linn. and its isolated constituent plumbagin: a review. Int J Pharm Sci Res. 2020;11(10):4859-65.

doi: 10.13040/IJPSR.0975-8232.11(10).4859-65.

- Zhao YL, Lu DP (). Effects of plumbagin on the human acute promyelocytic leukemia cells in vitro. Zhongguo Shi Yan Xue Ye Xue Za Zhi. 2006;14(2):208-11. PMID 16638181.
- 59. Sandur SK, Ichikawa H, Sethi G, Ahn KS, Aggarwal BB. Plumbagin (5-hydroxy-2-methyl-1,4-naphthoquinone) suppresses NF-kappaB activation and NF-kappaBregulated gene products through modulation of p65 and IkappaBalpha kinase activation, leading to potentiation of apoptosis induced by cytokine and chemotherapeutic agents. J Biol Chem. 2006;281(25):17023-33.

doi: 10.1074/jbc.M601595200, PMID 16624823.

60. Hiradeve S, Danao K, Kharabe V, Mendhe B. Evaluation of anticancer activity of Plumbago zeylanica Linn leaf extract. Int Jour of Biomed Res. 2010;1(2):01-9.

- doi: 10.7439/ijbr.v1i2.52.
 61. Li J, Li J, Cai G, Shen L, Lu F. Proapoptotic and growth-inhibitory effects of plumbagin on human gastric cancer cells via suppression of signal transducer and activator of transcription 3 and protein kinase B. Altern Ther Health Med. 2017;5382. Yadav P, S, Goutam MP. Anticancer efficacy of Plumbago zeylanica L. a review. Int J Sci Res Rev. 2019;8(7):290-307.
- 62. De paiva SR, Marques SD, Figueiredo MR, Kaplan MAC. plumbagin ales: A pharmacological approach. Floresta Ambiente. 2003;10(1):98-105.
- 63. Premakumari P, Rathinam K, Santhakumari G. Antifertility activity of plumbagin. Indian J Med Res. 1977;65(6):829-38. PMID 604259. Bhargava SK. Effects of plumbagin on reproductive function of male dog. Ind J Exp Biol. 1984;22:153-6. Saxena BP, Thappa RK, Tikku K, Sharma A, Suri OP. Effect of plumbagin on gonadotrophic cycle of the housefly Musca domestica. Indian J Exp Biol. 1996;34(8):739-44.
- 64. Dhar AK. Propagation of Plumbago zeylanica. J Med Aromat Plant Sci. 1999;21(2):304-7.
- Devarshi P, Patil S, Kanse A. Antiimplantation and abortive effects of Plumbago zeylanica in albino rats. Ind. J. Comp Ani Physiol. 1988;6(1):23-7.
- Chowdhury AK, Chakder SK, Khan AKA. Isolation and characterization of chemical constituents of Plumbago zeylanica roots. J Bangla Acad Sci. 1981;5(1):71-4.
- Azad Choudhary AK, Sushanta KC, Azadkhan AK. Antifertility activity of Plumbago zeylanica Linn. Root. Ind J Med Res. 1982;76:99-101.
- 68. Vishnukanta and Rana AC: evaluation of the antifertility activity of the hydroalcoholic extract of the Leaves of Plumbago zeylanica I. (plumbaginaceae) in female wistar rats. Indian J Pharm Educ Res. 2010 Jan–Mar;44(1):49-55.
- Simonsen HT, Nordskjold JB, Smitt UW, Nyman U, Palpu P, Joshi P, Varughese G. In vitro screening of Indian medicinal plants for antiplasmodial activity. J Ethnopharmacol. 2001;74(2):195-204. doi: 10.1016/S0378-8741(00)00369-X.
- Suraveratum N, Krungkrai SR, Leangaramgul P, Prapunwattana P, Krungkrai J. Purification and characterization of Plasmodium falciparum succinate dehydrogenase. Mol Biochem Parasitol. 2000;105(2):215-22. doi: 10.1016/s0166-6851(99)00180-2, PMID 10693744.
- Bachhawat A, Mohamed JS, Shihabudeen TK. Screening of fifteen Indian ayurvedic plants for alphaglucosidase inhibitory activity and enzyme kinetics. Int J Pharm Sci. 2009;3(4):267-74.
- 72. Jia Y, Jing J, Bai Y, Li Z, Liu L, Luo J, Liu M, Chen H. Amelioration of Experimental Autoimmune Encephalomyelitis by Plumbagin through Down-Regulation of JAK-STAT and NF-κB Signaling Pathways. PLOS ONE. 2011;6(10):1-7. doi: 10.1371/journal.pone.0027006.
- 73. Ram A. Effect of Plumbago zeylanica in hyperlipidaemic rabbits and its modification by vitamin E. Indian J Pharmacol. 1996;28(3):161-6.

- Kirtikar KR, Basu BD. Indian medicinal plants. Vol. II. Dehradun, India: Bishen Mahendra Pal Singh; 1993. p. 1466-8.
- Oyedapo OO. Studies on bioactivity of the root extract of Plumbago zeylanica. Int J Pharmacogn. 1996;34(5):365-9. doi: 10.1076/phbi.34.5.365.13249.

 Vishnukanta, Rana AC. Analgesic and antiinflammatory activity of hydroalcoholic extract of Plumbago zeylanica leaf extract. Pharmacogn Mag. 2008, Jul–Sep;15;Suppl:S133-136.

- Aleem M. Anti-Inflammatory and Anti-Microbial Potential of Plumbago zeylanica L.: A Review. J Drug Delivery Ther;10(5-s):229-35. doi: 10.22270/jddt.v10i5-s.4445.
- Chen YC, Tsai WJ, Wu MH, Lin LC, Kuo YC. Suberosin inhibits proliferation of human peripheral blood mononuclear cells through the modulation of the transcription factors NF-AT and NF-kappaB. Br J Pharmacol. 2007;150(3):298-312. doi: 10.1038/sj.bjp.0706987.
- Dai Y, Hou LF, Chan YP, Cheng L, but PPH. Inhibition of immediate allergic reactions by ethanol extract from Plumbago zeylanica Stems. Biol Pharm Bull. 2004;27(3):429-32. doi: 10.1248/bpb.27.429.
- 80. Amrutha N, Hemalatha T, Anjali M, Eswar Tony D, Nadendla R. Pharmacological evaluation of Plumbago zeylanica leaf extracts for anxiolytic activity by using open field test. J Harmon Res Pharm. 2016;5(1):7-10.
- Desta B. Ethiopian traditional herbal drugs. Part II: Antimicrobial activity of 63 medicinal plants. J Ethnopharmacol. 1993;39(2):129-39. doi: 10.1016/0378-8741(93)90028-4, PMID 8412246.
- Ahmad I, Mehmood Z, Mohammad F. Screening of some Indian medicinal plants for their antimicrobial properties. J Ethnopharmacol. 1998;62(2):183-93. doi: 10.1016/S0378-8741(98)00055-5.
- Beg AZ, Ahmad I. Beg AZ and Ahmadi, Effect of Plumbago zeylanica extract and certain curing agents on multidrug resistant bacteria of clinical origin. World J Microbiol Biotechnol. 2000;16(8/9):841-4. doi: 10.1023/A:1008991724288.
- Dama LB, Poul BN, Jadhav BV. Antimicrobial activity of plumbagin and related naphthoquinones. J. Ecol. Environ Monit. 1998;8(3-4):213.
- 85. Chopade BA, Patwardhan RB, Dhakephalkar PK. Acinetobacter infections in India: genetic and molecular biology studies and some approaches to the problem. In: Kumar S, Sen AK, Dutta GP, Sharma RN, editors CSIR Publications & Information Directorate Tropical diseases: molecular biology and control strategies. New Delhi, India; 1994. p. 704-17.
- Wang YC, Huang TL. Anti-Helicobacter pylori activity of Plumbago zeylanica L. FEMS Immunol Med Microbiol. 2005;43(3):407-12.

doi: 10.1016/j.femsim.2004.10.015, PMID 15708315.

- Devi CK, Krishna DG. Pharmacognostic, phytochemical and biological study of Plumbago zeylanica. Int JNat. Products Res. 2012;1(2):21-3.
- Banik B, Sarkar P, Sultana F, Saikia M, Dey A. In-vitro antimicrobial screening with phytochemical study of Plumbago zeylanica L. collected from two regions of Eastern Himalayas- A comparative study. Int J Phytopharmacol 2014. Vol. 4(5). p. 120-3.

- Mehmood Z, Ahmad I, Mohammad F, Ahmad S. Indian medicinal plants: A potential source for anticandidal drugs. Pharm Biol. 1999;37(3):237-42. doi: 10.1076/phbi.37.3.237.6296.
- Mahoney N, Molyneux RJ, Campbell BC. Regulation of aflatoxin production by naphthoquinones of walnut (Juglans regia). J Agric Food Chem. 2000;48(9):4418-21.

doi: 10.1021/jf0003449, PMID 10995372.

- Skinner FA. The antibiotics. In: Peach K, Tracy HV, editors, Published Springer-Verlag Modern methods of plant analysis. Vol. 3. Germany: West; 1993. p. 626-725.
- 92. Gebre-Mariam T, Neubert R, Schmidt PC, Wutzler P, Schmidtke M. Antiviral activities of some Ethiopian medicinal plants used for the treatment of dermatological disorders. Journal of Ethnopharmacology. 2006;104(1-2):182-7. doi: 10.1016/j.jep.2005.08.071.
- Fujii N, Yamashita Y, Arima Y, Nagashima M, Nakano H. Induction of topoisomerase II-mediated DNA cleavage by the plant naphthoquinones plumbagin and shikonin. Antimicrob Agents Chemother. 1992;36(12):2589-94.

doi: 10.1128/aac.36.12.2589, PMID 1336338.

- 94. Jeyachandran R, Mahesh A, Cindrella L, Sudhakar S, Pazhanichamy K. Antibacterial activity of plumbagin and root extracts of Plumbago zeylanica L. Acta Biol Cracoviensia S Bot. 2009;51(1):(1):17-22.
- Lakhmi VV, Padma S, Polasa H. Elimination of multidrug-resistant plasmid in bacteria by plumbagin, a compound derived from a plant. Curr Microbiol. 1987;16(3):159-61. doi: 10.1007/BF01568396.
- 96. Maniafu BM, Wilber L, Ndiege IO, Wanjala CC, Akenga TA. Larvicidal activity of extracts from three Plumbago spp against Anopheles gambiae. Mem Inst Oswaldo Cruz. 2009;104(6):813-7. doi: 10.1590/S0074-02762009000600002.
- 97. Patil CD, Patil SV, Salunke BK, Salunkhe RB. Bioefficacy of Plumbago zeylanica (Plumbaginaceae) and Cestrum nocturnum (Solanaceae) plant extracts against Aedes aegypti (Diptera: Culicide) and nontarget fish Poecilia reticulata. Parasitol Res. 2011;108(5):1253-63. dai: 10.1007/c00436.010.2174.6. PMID 21107959.

doi: 10.1007/s00436-010-2174-6, PMID 21107859.

- Rao JV, Sreenivasan C, Makkapati AK. Plumbagin effect on growth and metamorphosis of housefly Musca domestica L. (Diptera: Muscidae). Int Pest Control. 1996;38(1):24-5. > Article > pdf > floram-10-1-98.
- 99. Satyanarayana R, Gujar GT. Bioactivity of naturally occurring plumbaginoids against Dysdercus koenigii. Fabricius. 1999. Shashpa. Vol. 6(1). p. 29-35. Available from: http://www.floram.org. > Article > pdf > floram-10-1-98 [cited 7/11/2020].
- 100. Villavicencio MA, Perez-Escandon BE. Plumbagin activity (from Plumbago pulchella Boiss. Plumbaginaceae) as a feeding deterrent for three species of Orthopter. Folia Entomol Mex. 1992; 86:191-8.
- Krishnayya PV, Rao PJ. Effect of plumbagin on chitin, cuticular proteins, medium neurosecretory cells, and corpora allata of Helicoverpa armigera Hübner larvae.

Proceedings of the Indian Scientific Academy, part B. J Biol Sci. 1995;61(2):127-35.

102. Banerjee S, Magdum S, Kalena GP, Banerji A. Insect growth regulatory activity of naturally occurring quinones and their derivatives in Dysdercus koenigii Fabr. Hern Pyrrhocoridae, Appl Entomol.. 2001;125(1-2):25-30.

doi: 10.1111/j.1439-0418.2001. 00489.x.

- Ghosh D, Som K, Dinda B, Chel G. Potentiality of plumbagin to Culex fatigans. A growth inhibitor. J Adv Zool. 1994;15(2):112-5.
- Chockaligan S, Thenmozhi S. Undari MSN. Larvicidal activity of different products against mosquito larvae. J Environ Biol. 1990;11(2):101-4. Corpus ID: 82020644.
- Durga R, Sridhar P, Polasa H. Antimutagenic activity of plumbagin in Ames Salmonella typhimurium test. Indian J Med Res. 1992; 96:143-5. PMID 1428056.
- 106. Farr SB, Natvig DO, Kogoma T. Toxicity and mutagenicity of plumbagin and the induction of a possible new DNA repair pathway in Escherichia coli. J Bacteriol. 1985;164(3):1309-16. doi: 10.1128/JB.164.3.1309-1316.1985, PMID 2933393, PMCID PMC219331.
- Aqil F, Zahin M, Ahmad I. Antimutagenic activity of methanolic extracts of four ayurvedic medicinal plants. Indian J Exp Biol. 2008;46(9):668-72. PMID 18949897.
- 108. Bhavprakash, Vairabhaha SR, Chaukhambha Sanskrit Sansthan V. Vol I. 1997; p 70-1.
- Roy A, Bharadvaja N. A review on pharmaceutically important medical plant: Plumbago zeylanica. J ayu. Herb Med. 2017;3(4):225-8.
- 110. Mulke VG, Ghotankar AM. Therapeutic uses of Chitraka (Plumbago zeylanica linn.) with a note on its pharmacological actions -A review. World j. Pharm Res. 2020;6(4):56-9.
- III. Harborne JB. Comparative biochemistry of the flavonoids-IV. Phytochemistry. 1967;6(10):1415-28. doi: 10.1016/S0031-9422(00)82884-8.
- Itoigawa M, Takeya K, Furukawa H. Cardiotonic action of plumbagin on guinea-pig papillary muscle. Planta Med. 1991;57(4):317-9. doi: 10.1055/s-2006-960106, PMID 1775570.
- 113. Samhita C, Shree Gulab-kunverba Ayurvedic Society, Jamnagar SS. Adhyaya. 1946;4:63.
- 114. D'Astafort D. Plumbagin from P. zeylanica. J Pharm Sci. 1829;14:441.
- II5.
 Fieser LF, Dunn JT.
 Synthesis of plumbagin.
 J Am

 Chem
 Soc.
 1936;58(4):572-5.
 doi:

 10.1021/ja01295a010.
- 116. Imlay J, Fridovich I. Exogenous quinones directly inhibit the respiratory NADH dehydrogenase in Escherichia coli. Arch Biochem Biophys. 1992;296(1):337-46. doi: 10.1016/0003-9861(92)90581-G.
- 117. Demma J, Engidawork E, Hellman B. Potential genotoxicity of plant extracts used in Ethiopian traditional medicine. J Ethnopharmacol. 2009;122(1):136-42. doi: 10.1016/j.jep.2008.12.013.
- 118. Krishna PV, Rao PJ. Plumbagin a natural bioactive principle against insect. The Andra Agric. 1996;43:1-5.
- 119. Sharma A, Singh RT, Sehgal V, Handa SS. Antihepatotoxic activity of some plants used in herbal formulations. Fitoterapia. 1991;62(2):131-8.

- 120. Abdul KM, Ramchender RP. Modulatory effect of plumbagin (5-hydroxy-2-methyl-1,4-naphthoquinone) on macrophage functions in BALB/c mice. I. Potentiation of macrophage bactericidal activity. Immunopharmacology. 1995;30(3):231-6. doi: 10.1016/0162-3109(95)00027-Q.
- 121. Mossa JS, El-Feraly FS, Muhammad I. Antimycobacterial constituents fromJuniperus procera, Ferula communis andPlumbago zeylanica and theirin vitro synergistic activity with isonicotinic acid hydrazide. Phytother Res. 2004;18(11):934-7. doi: 10.1002/ptr.1420.
- Molnár J. Antiplasmid activity of tricyclic compounds. Methods Find Exp Clin Pharmacol. 1988;10(7):467-74. PMID 3047509.
- 123. Padhye SB, Kumbhar AS, Chopade BA. Reversal of drug resistance by plumbagin (5-hydroxy-2methyl-1-4napthoquinione) in microorganisms. In: Indo-French Symposium on Apoptosis and Multiple Drug Resistance; 1997. p. 45-6.
- 124. Patil AP, Mukherjee AR, Kumbhar AS, Padhye SB, Chopade BA. DNA strand session and plasmid curing activity of an Indian folk medicine constituent Chitrak. In:. In: Recent Advances in Chemotherapy. Proceedings of the 18th international congress of chemotherapy. Washington, DC: American Society for Microbiology; 1994. p. 504-5.
- 125. Patwardhan RB, Dhakephalkar PK, Chopade BA. Antibacterial and plasmid curing activities of root extracts of Plumbago zeylanica. Int J Herbo Med. 2015;2(1):13-25.
- Deshpande LM, Chopade BA. Plasmid mediated silver resistance in Acinetobacter baumannii. BioMetals. 1994;7(1):49-56.
 - doi: 10.1007/BF00205194.
- 127. Chopade BA, Patwardhan RB, Vaidya VC, Khairnar S, Dhakephalkar PK, Padhye SB. Curing of antibiotic and metal resistance plasmids in Acinetobacter baumannii. In: Kumar S, Sen AK, Dutta GP, Sharma RN, editors Tropical diseases: molecular biology and control

strategies. CSIR Publications & Information Directorate, New Delhi. India:; 1994. p. 695-703.

- 128. Chinchalkar LJ, Kapadnis BP, Chopade BA. Novel plasmid mediated aminoglycoside resistance in Acinetobacter baumannii BL110. In: Proceedings of the DAE symposium on molecular biology of microorganisms. Mumbai, India: Department of Atomic Energy, Government of India; 1992. p. 56-61.
- 129. Dhakephalkar PK, Bhamare SA, Kunte DP, Pashine A, Purnapatre K, Chopade BA. Mobilization of nonconjugative plasmids in Acinetobacter baumannii C11 by P incompatibility group plasmid RP4. In: Proceedings of the DAE symposium on molecular biology of microorganisms. Mumbai, India: Department of Atomic Energy, Government of India; 1992. p. 78-81.
- 130. Dhakephalkar PK, Murthi S, Sheshadri RG, Puntambekar M, Chopade BA. Genetic studies on pUPI102: plasmid encoding resistance to tetracycline, gentamycin, neomycin and mercury in Acinetobacter junii ACN4. In: Proceedings of the DAE symposium on molecular biology of microorganisms. Mumbai, India: Department of Atomic Energy, Government of India; 1992. p. 72-6.
- 131. Patwardhan RB. Studies on human pathogenic Acinetobacter species [masters Phil thesis] Submitted to University of Pune. India; 1990.
- 132. Shakibaie MR, Dhakephalkar PK, Kapadnis BP, Chopade BA. Removal of silver from photographic wastewater effluent using Acinetobacter baumannii BL54. Can J Microbiol. 1999;45:995-1000. doi: 10.1139/w99-077.
- 133. Shakibai MR, Dhakephalkar PK, Kapadnis BP, Chopade BA. Silver resistance in Acinetobacter baumannii BL54 occurs through binding to Ag- binding protein. Iran J Biotechnol. 2003;1(1):41-6.
- Bharathi A, Polasa H. Elimination of ColE I (pBR322 and pBR329) plasmids in Escherichia coli by αsantonin. FEMS Microbiol Lett. 1990;68(1-2):213-5. doi: 10.1111/j.1574-6968.1990.tb04151.x.